

## The Effect of Plant Growth Regulators on Shoot Regeneration of Two Wild Almond Species *Amygdalus scoparia* L. and *Amygdalus lycioides* Spach.

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### ABSTRACT

Two species of *Amygdalus scoparia* L. and *A. lycioides* Spach. from the Rosaceae family can be found in suitable environmental conditions due to their high resistance to drought and heat and resistance to calcareous soils and attacking nematodes and diseases. The propagation of fruit tree rootstocks through vegetative means can be enhanced through the optimization of micropropagation techniques. This research was done in plant tissue culture laboratory of the Agricultural and Natural Resources Research Center of Kermanshah in 2018. In this research, the improved micropropagation protocol of these plants, explants of disinfected nodes in MS medium in combination with BA (6-Benzyladenine) hormones, at 6 levels (0, 0.25, 0.5, 1, 2, and 3 mg L<sup>-1</sup>) in four levels of Indole Butyric Acid (0, 0.1, 0.25 and 0.5 mg L<sup>-1</sup>). The experiment was carried out with four replications in a completely randomized factorial design. The results indicated that the highest percentage of explant regeneration in the *A. lycioides* genotype was achieved with a combination of 0.5 mg L<sup>-1</sup> BA and 0.25 mg L<sup>-1</sup> IBA, while in the *A. scoparia* genotype, the highest regeneration percentage was observed with a combination of 2 mg L<sup>-1</sup> BA and 0 mg L<sup>-1</sup> IBA. The *A. lycioides* genotype exhibited the highest number of shoots when treated with 0.5 mg L<sup>-1</sup> BA and 0.25 mg L<sup>-1</sup> IBA, while the *A. scoparia* genotype showed the maximum shoot number at a BA concentration of 2 mg L<sup>-1</sup> and IBA concentration of 0 mg L<sup>-1</sup>.

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### 1. Introduction

Iran is the origin of most wild almond species in the world and about 21 species have been reported in different regions of the country (Rahemi *et al.*, 2011). It represents a wide range of morphological characteristics that have grown naturally in different regions of the country. Due to their adaptability to extreme environmental conditions (drought), these can be used as rootstocks in almond or peach (Rahemi *et al.*, 2011; Ansari *et al.*, 2019; Imani *et al.*, 2021). *Amygdalus lycioides* is a native species of Iran. It is a small deciduous shrub with tough and thorny branches that height is about 1 to 1.5 meters (Rahemi and Gradziel, 2024). *Amygdalus scoparia* is another native species of Iran that exists in many areas of the country.

It is a large deciduous shrub with a height of about 2 to 2.5 meters, which can sometimes reach up to 6 meters (Fig. 1). It produces a large number of long, straight, narrow, smooth green shoots (Rouhi *et al.*, 2007).

Drought stress is one of the most important limitations in the world that restricts the yield of agricultural products (Alaei, 2019). In order to prevent the reduction of the yield of agricultural products in drought conditions, it is necessary to choose plants that tolerate unfavorable environmental conditions (Ashrafi *et al.*, 2018). So, one of the ways to reduce drought stress is the selection of drought-tolerant rootstocks. Resistance to drought stress in plants is caused by the interaction of biochemical, physiological and morphological characteristics (Lotfi *et al.*, 2019).

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The species *A. scoparia* and *A. lycioides* are the most important species that are used as almond rootstocks in non-irrigated conditions. Development of drought-tolerant almond orchards using *A. scoparia* rootstocks will result in more sustainable production in arid regions (Khadivi-Khub and Anjam, 2016). Phenotypic diversity is the essential characteristic of plants to respond to environmental changes for their long-term survival (Imani et al., 2021). Some reports showed that leaves of *A. scoparia* species fall under stress conditions, but some leaves of *A. lycioides* species survive and are able to carry out photosynthesis (Rouhi et al., 2007). Also, these species grow in shallow and dry limestone soils (Ali et al., 2015).

Climate change is one of the most important agricultural challenges in the world, especially in dry and semi-arid climates. To deal with this problem, it will be necessary to introduce new varieties and rootstocks (Gharaghani et al., 2017). Achieving dwarf rootstocks resistant to pests, diseases and environmental stress is one of the goals of many rootstocks breeding programs (Panis et al., 2020; Volk et al., 2023). It has been reported that grafting almond cultivars on *A. scoparia* causes a decrease in plant height and an increase in almond fruit yield under drought stress conditions and calcareous soils. Almond cultivars are propagated sexually and asexually (Rahemi et al., 2011). On the other hand, rooting in almonds is difficult and propagation through seeds leads to genetic variation and will not be suitable for use as a clone. Therefore, the propagation method through tissue culture is an efficient alternative system (Farsi et al., 2016; Eshghi Khas et al., 2020).

In order to use a specific species as a vegetative rootstock and propagate it through tissue culture, it is necessary to develop effective regeneration and propagation. So this research reports a micropropagation protocol using segment nodal explants of *A. lycioides* and *A. scoparia* species in Iran. For commercial propagation of these species in the nursery, an optimal protocol is required for micropropagation of this species. We did not find any reports on shoot micropropagation of *A. lycioides*. Therefore this is the first study reporting the *in vitro* regeneration of *A. lycioides*. Therefore, in this research,

we investigated the micropropagation of *A. lycioides* and *A. scoparia* species using different concentrations of cytokinin and auxin.

## 2. Materials and methods

This research was done in the Plant Tissue Culture Laboratory of the Agricultural and Natural Resources Research Center of Kermanshah in 2018. The growing branches were collected in May from the young parts of two wild almond species of *A. scoparia* and *A. lycioides* in the Bilawar region of Kermanshah province in Iran (Fig. 1).

The branches were kept in an ice box during transport to the laboratory. The shoot samples were washed under tap water for 30 minutes to remove surface contamination. Surface sterilization was performed inside the laminar air flow cabinet by immersing the explants in 70% (v/v) ethanol for 30 seconds and by immersing them in 1% (v/v) sodium hypochlorite solution for 15 minutes. After washing three times in sterile double distilled water, sterilized uniform nodal branch segments (length 3-4 cm) were cultured on MS culture medium in combination with BA (benzyladenine) hormones, at 6 levels (0, 0.25, 0.5, 1, 2, and 3 mg L<sup>-1</sup>) in four levels of Indole Buteric Acid (0, 0.1, 0.25 and 0.5 mg L<sup>-1</sup>). The pH of the medium was adjusted to 5.8 using 0.1N of NaOH or HCl and before autoclaving, the medium was solidified by adding 0.8% agar.

For regeneration of shoot culture, individual sterile nodal segments were placed in 40 ML polycarbonate tubes containing medium. The cultures were incubated in a growth chamber under 16/8 h light/dark cycle under 40 mol m<sup>-2</sup> s<sup>-1</sup> illuminations in cool, white fluorescent light at 25 ± 1°C. After 6 weeks, regeneration percentage, shoot number, shoot length, leaf number, leaf length, and internodal distance were measured.

The experiment was carried out as factorial in a completely randomized design with four replications. Factorial analysis of variance was conducted using statistical analysis system (SAS) and all data were analyzed according to Duncan's Multiple Range Test ( $p \leq 0.05$ ).

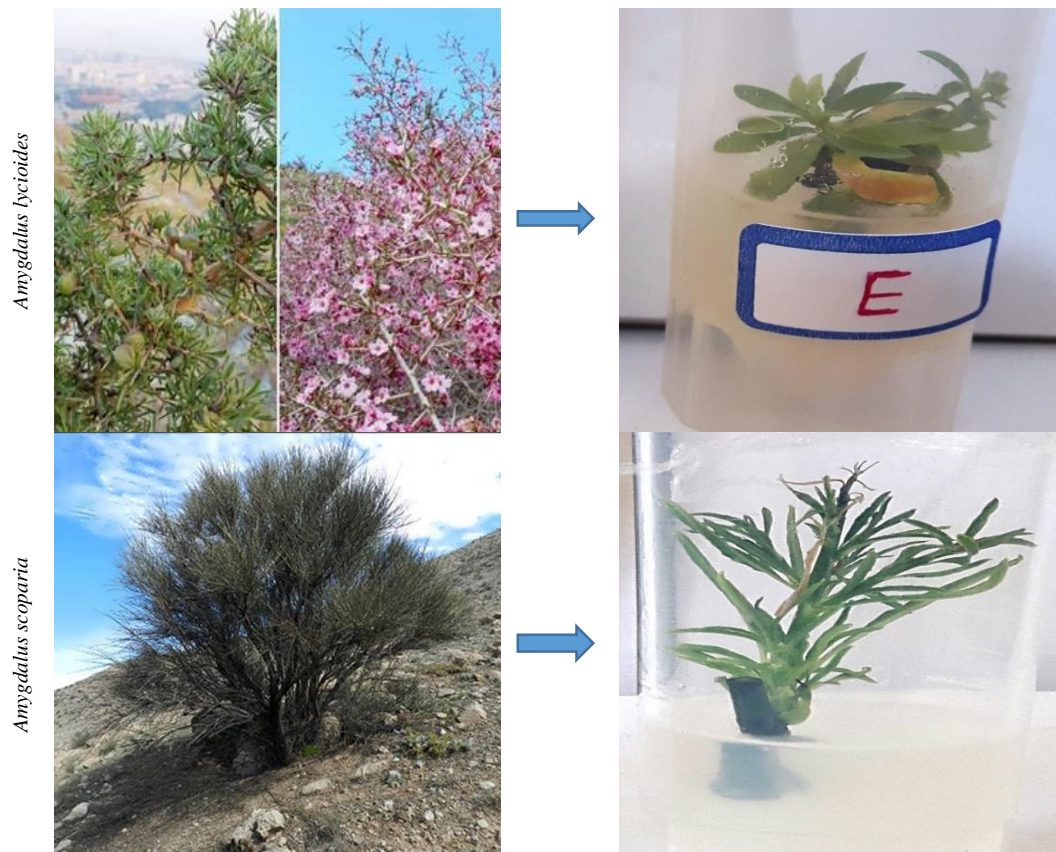


Figure 1. Shoot proliferation: *Amygdalus lycioides* (Combination of 0.5 mg L<sup>-1</sup> BA and 0.25 mg L<sup>-1</sup> IBA). *Amygdalus scoparia* (Combination of 2 mg L<sup>-1</sup> BA and 0 mg L<sup>-1</sup> IBA).

### 3. Results and discussion

For regeneration of wild almond species including *A. scoparia* and *A. lycioides*, individual sterile nodal segments were cultured on MS medium supplemented with BA at the concentration of 0, 0.25, 0.5, 1, 2, and 3 mg L<sup>-1</sup>, or BA combined with IBA at the concentration of 0, 0.1, 0.25 and 0.5 mg L<sup>-1</sup>. Analysis of variance showed that the effect of different levels of genotype, growth regulators of cytokinin (BA) and auxin (IBA) and interaction effects between these factors on the regeneration percentage, shoot number, shoot length, leaves number, leaf length, and internodal distance traits were significant at ( $p \leq 0.01$ ) level.

#### 3.1. Regeneration percentage

The average comparison results showed that the lowest regeneration frequency was obtained on a hormone-free MS medium for both species. The BA concentrations less than 0.05 mg L<sup>-1</sup> did not show shoot induction and proliferation in both genotypes. The highest percentage of regeneration of explants in *A. lycioides* was obtained in the combination of 0.5 mg L<sup>-1</sup> BA and 0.25 mg L<sup>-1</sup> IBA. In the *A. scoparia* plant, the highest percentage of explant regeneration was

obtained in the combination of 2 mg L<sup>-1</sup> BA and 0 mg L<sup>-1</sup> IBA. In fact, the highest regeneration frequencies (98.25% and 93.00%) were obtained on MS medium supplemented with BAP and IBA for *A. scoparia* and *A. lycioides* genotypes, respectively (Table 1).

#### 3.2. Shoot number

Among BA concentrations, only 0.5, 1, 2, and 3 mg L<sup>-1</sup> treatments generated shoot, while there were no shoots in the media containing 0 and 0.25 mg L<sup>-1</sup> BA concentrations. Results revealed that *in vitro* shoot proliferation response increased by combining 0.5 mg L<sup>-1</sup> of BA with 0.25 mg L<sup>-1</sup> of IBA in *A. lycioides* genotype. However, the highest number of shoots per explant of *A. scoparia* genotype was recorded only with cultivated in 1 mg L<sup>-1</sup> BA medium without the use of IBA. On this medium, *A. lycioides* and *A. scoparia* explants produced 2.12 and 4.25 average number of shoots per explant, respectively (Table 1).

#### 3.3. Shoot length

The results showed that the maximum length of shoots in *A. lycioides* plant was obtained in the combination of 1 mg L<sup>-1</sup> BA and 0.1 mg L<sup>-1</sup> IBA. In *A.*



*scoparia* plant, the maximum shoot length was observed in the combination of 2 mg L<sup>-1</sup> BA and 0 mg L<sup>-1</sup> IBA. Culturing the explants on various media showed different results in terms of shoot length. Culturing on this medium, *A. lycioides* and *A. scoparia* explants generated 4.37 and 23.75 cm as average length of the tallest shoot per explant, respectively (Table 1).

**Table 1. Mean comparison of the effect of different BA and IBA concentrations on on different traits of two wild almond genotypes.**

plant growth regulator (mg L <sup>-1</sup> )	Regeneration (%)	Shoot number		Shoot length (cm)			
		wild almond genotypes	wild almond genotypes	wild almond genotypes	wild almond genotypes		
BA	IBA	<i>A. lycioides</i>	<i>A. scoparia</i>	<i>A. lycioides</i>	<i>A. scoparia</i>	<i>A. lycioides</i>	<i>A. scoparia</i>
0	0	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.1	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.25	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.5	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
0.25	0	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.1	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.25	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.5	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
0.5	0	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.1	31.00 <sup>h</sup>	53.50 <sup>e</sup>	0.48 <sup>h</sup>	0.48 <sup>h</sup>	3.20 <sup>jk</sup>	3.25 <sup>j</sup>
	0.25	93.00 <sup>b</sup>	77.25 <sup>c</sup>	2.12 <sup>c</sup>	0.71 <sup>g</sup>	2.75 <sup>kl</sup>	7.43 <sup>c</sup>
	0.5	26.25 <sup>i</sup>	73.25 <sup>d</sup>	1.00 <sup>f</sup>	0.63 <sup>g</sup>	2.37 <sup>l</sup>	3.25 <sup>j</sup>
1	0	0.00 <sup>k</sup>	98.25 <sup>a</sup>	0.00 <sup>j</sup>	4.25 <sup>a</sup>	0.00 <sup>m</sup>	10.25 <sup>b</sup>
	0.1	48.75 <sup>f</sup>	73.75 <sup>d</sup>	1.37 <sup>he</sup>	2.11 <sup>c</sup>	4.37 <sup>g</sup>	4.81 <sup>f</sup>
	0.25	0.00 <sup>k</sup>	48.25 <sup>f</sup>	0.00 <sup>j</sup>	1.00 <sup>f</sup>	0.00 <sup>m</sup>	6.25 <sup>de</sup>
	0.5	0.00 <sup>k</sup>	0.00 <sup>k</sup>	0.00 <sup>j</sup>	0.00 <sup>j</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
2	0	0.00 <sup>k</sup>	22.00 <sup>j</sup>	0.00 <sup>j</sup>	0.44 <sup>h</sup>	0.00 <sup>m</sup>	23.75 <sup>a</sup>
	0.1	43.75 <sup>g</sup>	72.00 <sup>d</sup>	1.43 <sup>e</sup>	1.94 <sup>d</sup>	4.17 <sup>gh</sup>	4.25 <sup>g</sup>
	0.25	0.00 <sup>k</sup>	96.25 <sup>a</sup>	0.00 <sup>j</sup>	2.42 <sup>b</sup>	0.00 <sup>m</sup>	3.75 <sup>hi</sup>
	0.5	0.00 <sup>k</sup>	45.25 <sup>g</sup>	0.00 <sup>j</sup>	0.53 <sup>h</sup>	0.00 <sup>m</sup>	3.37 <sup>ij</sup>
3	0	0.00 <sup>k</sup>	25.00 <sup>ij</sup>	0.00 <sup>j</sup>	0.47 <sup>h</sup>	0.00 <sup>m</sup>	2.90 <sup>jk</sup>
	0.1	0.00 <sup>k</sup>	50.00 <sup>f</sup>	0.00 <sup>j</sup>	0.72 <sup>g</sup>	0.00 <sup>m</sup>	4.92 <sup>f</sup>
	0.25	0.00 <sup>k</sup>	73.75 <sup>d</sup>	0.00 <sup>j</sup>	1.46 <sup>e</sup>	0.00 <sup>m</sup>	6.50 <sup>d</sup>
	0.5	0.00 <sup>k</sup>	23.00 <sup>j</sup>	0.00 <sup>j</sup>	0.23 <sup>i</sup>	0.00 <sup>m</sup>	5.89 <sup>e</sup>

The means with the same letters in each trait are not significantly different at the 5% level using Duncan's multiple range tests.

### 3.4. Internode length

The results of comparing the averages showed that the maximum internode distance of *A. lycioides* plant was obtained in MS culture medium with 0.5 mg L<sup>-1</sup> BA and 0.1 mg L<sup>-1</sup> IBA. In *A. scoparia* plant, the highest internode distance in the environment was obtained with the combination of 1 mg L<sup>-1</sup> BA and 0.25 mg L<sup>-1</sup> IBA (Table 2).

### 3.5. Leaf number

As shown in Table 2, the number of leaves on shoots produced on media containing only 0.5, 1, 2, and 3 mg

L<sup>-1</sup> while there were no shoots in the media containing other BA concentrations. The highest number of leaves of *A. lycioides* plant was obtained in MS culture medium with 2 mg L<sup>-1</sup> of BA and 0.1 mg L<sup>-1</sup> of IBA. In *A. scoparia* plant, the highest number of leaves was obtained in the environment with the combination of 2 mg L<sup>-1</sup> BA and 0 mg L<sup>-1</sup> IBA (Table 2).

### 3.6. Leaf length

The results of comparing the averages showed that the maximum leaf length of *A. lycioides* plant was recorded in MS culture medium with 0.5 mg L<sup>-1</sup> BA and 0.5 mg L<sup>-1</sup> IBA. In the *A. scoparia* plant, the maximum leaf length was obtained in the MS culture medium with 0.5 mg L<sup>-1</sup> BA and 0 mg L<sup>-1</sup> IBA (Table 2).

**Table 2. Mean comparison of the effect of different BA and IBA concentrations on on different traits of two wild almond genotypes.**

plant growth regulator (mg L <sup>-1</sup> )	Leaf number	Leaf length		Internode length			
		wild almond genotypes	wild almond genotypes	wild almond genotypes	wild almond genotypes		
BA	IBA	<i>A. lycioides</i>	<i>A. scoparia</i>	<i>A. lycioides</i>	<i>A. scoparia</i>	<i>A. lycioides</i>	<i>A. scoparia</i>
0	0	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.1	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.25	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.5	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
0.25	0	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.1	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.25	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.5	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
0.5	0	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
	0.1	7.18 <sup>e</sup>	7.75 <sup>e</sup>	2.97 <sup>gh</sup>	3.55 <sup>ef</sup>	0.44 <sup>i</sup>	0.42 <sup>ij</sup>
	0.25	13.50 <sup>b</sup>	14.00 <sup>b</sup>	3.00 <sup>gh</sup>	3.56 <sup>fe</sup>	0.21 <sup>l</sup>	0.53 <sup>gh</sup>
	0.5	11.25 <sup>c</sup>	6.50 <sup>f</sup>	3.81 <sup>e</sup>	2.93 <sup>ghi</sup>	0.21 <sup>l</sup>	0.51 <sup>h</sup>
1	0	0.00 <sup>i</sup>	13.88 <sup>b</sup>	0.00 <sup>l</sup>	8.81 <sup>a</sup>	0.00 <sup>m</sup>	0.73 <sup>c</sup>
	0.1	11.50 <sup>c</sup>	7.15 <sup>e</sup>	3.25 <sup>fg</sup>	4.32 <sup>d</sup>	0.38 <sup>j</sup>	0.68 <sup>d</sup>
	0.25	0.00 <sup>i</sup>	4.87 <sup>g</sup>	0.00 <sup>l</sup>	3.85 <sup>e</sup>	0.00 <sup>m</sup>	1.27 <sup>a</sup>
	0.5	0.00 <sup>i</sup>	0.00 <sup>i</sup>	0.00 <sup>l</sup>	0.00 <sup>l</sup>	0.00 <sup>m</sup>	0.00 <sup>m</sup>
2	0	0.00 <sup>i</sup>	39.50 <sup>a</sup>	0.00 <sup>l</sup>	2.87 <sup>hi</sup>	0.00 <sup>m</sup>	0.60 <sup>ef</sup>
	0.1	14.13 <sup>b</sup>	7.50 <sup>e</sup>	2.62 <sup>lj</sup>	5.62 <sup>b</sup>	0.29 <sup>k</sup>	0.56 <sup>fg</sup>
	0.25	0.00 <sup>i</sup>	6.50 <sup>f</sup>	0.00 <sup>l</sup>	3.87 <sup>e</sup>	0.00 <sup>m</sup>	0.62 <sup>e</sup>
	0.5	0.00 <sup>i</sup>	6.15 <sup>f</sup>	0.00 <sup>l</sup>	2.40 <sup>jk</sup>	0.00 <sup>m</sup>	0.55 <sup>gh</sup>
3	0	0.00 <sup>i</sup>	4.25 <sup>h</sup>	0.00 <sup>l</sup>	2.25 <sup>k</sup>	0.00 <sup>m</sup>	0.68 <sup>d</sup>
	0.1	0.00 <sup>i</sup>	5.25 <sup>g</sup>	0.00 <sup>l</sup>	3.25 <sup>fg</sup>	0.00 <sup>m</sup>	0.94 <sup>b</sup>
	0.25	0.00 <sup>i</sup>	10.50 <sup>d</sup>	0.00 <sup>l</sup>	4.75 <sup>c</sup>	0.00 <sup>m</sup>	0.62 <sup>e</sup>
	0.5	0.00 <sup>i</sup>	6.25 <sup>f</sup>	0.00 <sup>l</sup>	3.45 <sup>f</sup>	0.00 <sup>m</sup>	0.94 <sup>b</sup>

The means with the same letters in each trait are not significantly different at the 5% level using Duncan's multiple range tests.

As a general conclusion, it can be emphasized that different wild almond genotypes show different reactions to the type and concentration of plant growth regulators used in culture media. Therefore, it is

recommended that if a wild almond genotype is selected as a rootstock for almonds and if there is a need for micropropagation, it is recommended to use a special culture medium which is the best result for its multiplication.

The data obtained in this study indicated that the response of regeneration percentage of explants was greatly dependent on the concentration of BA combination with different concentrations of IBA. BA combination with the low concentration of IBA improves regeneration percentage of explants. The BA concentrations less than  $0.05 \text{ mg L}^{-1}$  did not show shoot induction and proliferation in both genotypes. The lowest value of regeneration was for all concentrations of BA combination with the high concentration of IBA combination ( $0.5 \text{ mg L}^{-1}$ ). Callus induction was observed within three weeks after inoculation of nodal segments on MS medium containing different concentrations of BA combination with the high concentration of IBA (Table 1). These results showed that the use of BA is necessary for the proliferation of explants. The proliferation of shoots in tissue culture conditions is influenced by many factors such as the type and concentration of plant growth regulators, explant type and genotypes which was consistent with the results of other researchers (Kodad et al., 2021). Both auxin and cytokinin hormones are important in explant organogenesis. Mozafari et al. (2016) reported that a low concentration of benzyl adenine in medium, decreased the number of shoots, but with an increase in BA concentration, the number of shoots increased. Various studies have shown that plant growth regulators are necessary for the regeneration of explants (Lizarraga et al., 2017).

MS media supplemented with BA, had a significant effect on shoot number. The results showed that a hormone-free MS nutrient medium does not shoot proliferation for both genotypes. A genotype effect was observed between the different tested genotypes since *A. lycioides* and *A. scoparia* show different proliferation rates in the same media. Regarding the interaction between genotype and plant growth regulators,  $0.5$  and  $1 \text{ mg L}^{-1}$  BA allowed for better results in both genotypes since proliferation rate was much more enhanced in MS medium than in  $2$  and  $3 \text{ mg L}^{-1}$  BA medium. *A. lycioides* exhibited significantly greater shoot numbers compared to most other treatments when  $0.25 \text{ mg L}^{-1}$  of IBA was combined

with  $0.5 \text{ mg L}^{-1}$  of BA. In contrast, the highest number of shoots in the *A. scoparia* genotype was achieved only with  $1 \text{ mg L}^{-1}$  of BA. BAP growth regulator is commonly used as shoot proliferation hormone in plant tissue culture. Renu and Singh (2018) also reported that the highest number of branches was observed in Fuji red apple on MS culture medium with  $4.44 \text{ M BAP}$  and  $0.05 \text{ NAA}$ . It is reported that in MS culture medium with  $0.5 \text{ mg L}^{-1}$  BAP and  $0.5 \text{ mg L}^{-1}$  GA<sub>3</sub>, causes 70% regeneration of Gisela-5 cherry rootstock explants (Thakur et al., 2016). In a study, it was found that culturing the shoot tips of the peach rootstock Hansen 536 on WPM medium +  $1 \text{ mg L}^{-1}$  BA and  $0.1 \text{ mg L}^{-1}$  NAA obtained the highest shoot regeneration (Sabbadini et al., 2019). The use of BA for shoot regeneration has also been reported in wild almond (Abbasi et al., 2019; Ebrahimi et al., 2022). Other researchers also reported that BA for shoot regeneration is more effective than other cytokinins (Nas et al., 2010).

Plant growth regulators had a great influence on shoot length. The results of this test showed that shoot length of genotype was affected by BA and IBA combinations. The two genotypes significantly were different in shoot length as affected by the treatments. The *A. scoparia* genotype produced the longest shoot length, while the shortest value was recorded with the *A. lycioides*. Our experimental results are in agreement with those observed for the Peach Rootstocks (Abou Elyazid Doaa et al., 2021) and *A. scoparia* (Abbasi et al., 2019). It has been reported that the most effective medium for increasing shoot length is MS medium containing  $0.5 \text{ mg L}^{-1}$  BA (Ak et al., 2021). This study showed that by increasing BA concentration to  $2 \text{ mg L}^{-1}$ , the number of leaves increased. In the almond genotype 2-22, the hormonal combination of  $1 \text{ mg L}^{-1}$  BA +  $0.05 \text{ mg L}^{-1}$  IBA resulted in the highest number of leaves and shoot length (Alizadeh-Arimi et al., 2020). Furthermore, according to Abbasi et al. (2019) for micropropagation of *A. scoparia* only  $2 \text{ mg L}^{-1}$  BA was the optimum concentration.

#### 4. Conclusion

Shoots regeneration in this study was influenced by the type and concentration of plant growth regulators and genotypes. The data obtained in this study indicated that the response of regeneration percentage of explants were greatly dependent on the

concentration of BA combination with different concentrations of IBA. BA combination with the low concentration of IBA improves regeneration percentage of explants.

### Conflict of interests

All authors declare no conflict of interest.

### Ethics approval and consent to participate

No humans or animals were used in the present research. The authors have adhered to ethical standards, including avoiding plagiarism, data fabrication, and double publication.

### Consent for publications

All authors read and approved the final manuscript for publication.

### Availability of data and material

All the data are embedded in the manuscript.

### Authors' contributions

All authors had an equal role in study design, work, statistical analysis and manuscript writing.

### Informed consent

The authors declare not to use any patients in this research.

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### References

- Abbasi F., Khadivi A., Taghizadeh M., ValizadehKaji B. 2019. Micropropagation of *Prunus scoparia*, a suitable rootstock for almond under drought conditions. *International Journal of Fruit Science* 19(2): 221-230. <https://doi.org/10.1080/15538362.2018.1539695>
- Abou Elyazid Doaa M., Gawish M.S., Eliwa G.I. 2021. Optimized protocol for micropropagation of cadaman and garnem peach rootstocks. *Journal of Plant Production* 12(7): 731-735. <https://doi.org/10.21608/jpp.2021.83432.1038>
- Ak B.E., Kiyar P.K., Hatipoglu I.H., Dikmetas B. 2021. Effects of different BA and IBA concentrations on proliferation and rooting of 'GARNEM' rootstock *in vitro* propagation. *International Journal of Agriculture, Environment and Food Sciences* 5(4): 470-476. <https://doi.org/10.31015/jaefs.2021.4.6>
- Alaei S. 2019. Essential oil content and composition of *Dracocephalum moldavica* under different irrigation regimes. *International Journal of Horticultural Science and Technology* 6(2): 167-175. <https://doi.org/10.22059/ijhst.2019.280572.294>
- Ali J.J.M., Nori I.M., Hama S.J., Rashed S.O. 2015. Water harvesting through utilization of wild almond as rootstocks for production of peach, apricot and plum under dry land farming in Sulaymaniyah region. *International Journal of Innovative Science, Engineering & Technology* 2(8): 705-724.
- Alizadeh-Arimi F., Yadollahi A., Imani A., Fakoor-Aryan M. 2020. Optimization of the sterilization and establishment steps for almonds 2-22 genotype. *Journal of Nuts* 11(4): 279-290. <https://doi.org/10.22034/jon.2020.1908563.1092>
- Ansari A., Gharaghani A. 2019. A comparative study of genetic diversity, heritability and inter-relationships of tree and nut attributes between *Prunus scoparia* and *P. elaeagnifolia* using multivariate statistical analysis. *International Journal of Horticultural Science and Technology* 6(1): 137-150. <https://doi.org/10.22059/ijhst.2019.276425.282>
- Ashrafi M., Azimi Moqadam M.R., Moradi P., Shekari F., Mohseni Fard E. 2018. Identification of drought tolerant and sensitive species of thyme through some physiological criteria. *International Journal of Horticultural Science and Technology* 5(1): 53-63. <https://doi.org/10.22059/ijhst.2018.255144.231>
- Ebrahimi M., Habashi A.A., Emadpour M., Kazemi N. 2022. Recovery of virusfree almond (*Prunus dulcis*) cultivars by somatic embryogenesis from meristem undergone thermotherapy. *Scientific Reports* 12: 14948. <https://doi.org/10.1038/s41598-022-19269-3>
- Eshghi Khas M., Abbasifar A., Valizadeh Kaji B. 2020. Optimization of *in vitro* propagation of purple passion fruit (*Passiflora edulis*), an important medicinal and ornamental plant. *International Journal of Horticultural Science and Technology* 7(3): 305-314. <https://doi.org/10.22059/ijhst.2020.297194.342>
- Farsi M., Fatahi Moghadam M.R., Zamani Z., Hassani D., Ahmadi A. 2016. The histology of minigrafting of Persian walnut trees cv. chandler. *International Journal of Horticultural Science and Technology* 3(2): 167-177. <https://doi.org/10.22059/ijhst.2016.62916>
- Gharaghani A., Solhjoo S., Oraguzie N. 2017. A review of genetic resources of almonds and stone fruits (*Prunus* spp.) in Iran. *Genetic Resources and Crop Evolution* 64: 611-640. <https://doi.org/10.1007/s10722-016-0485-x>
- Imani A., Amani G., Shamili M., Mousavi A., Hamed R., Rasouli M., José Martínez-García P. 2021. Diversity and broad sense heritability of phenotypic characteristic in almond cultivars and genotypes. *International Journal of Horticultural Science and Technology* 8(3): 281-289. <https://doi.org/10.22059/ijhst.2020.284452.303>
- Khadivi-Khub A., Sarooghi F., Abbasi F. 2016. Phenotypic variation of *Prunus scoparia* germplasm: Implications for

- breeding. *Scientia Horticulturae* 207: 193-202. <https://doi.org/10.1016/j.scienta.2016.05.023>
- Kodad S., Melhaoui R., Hano C., Addi M., Sahib N., Elamrani A., Abid M., Mihamou A. 2021. Effect of culture media and plant growth regulators on shoot proliferation and rooting of internode explants from Moroccan native Almond (*Prunus dulcis* Mill.) genotypes. *International Journal of Agronomy* 2021(1): 9931574. <https://doi.org/10.1155/2021/9931574>
- Lizarraga A., Fraga M., Ascasibar J., Gonzalez M.L. 2017. *In vitro* propagation and recovery of eight apple and two pear cultivars held in a germplasm bank. *American Journal of Plant Sciences* 8(9): 2238-2254. <https://doi.org/10.4236/ajps.2017.89150>
- Lotfi N., Soleimani A., Vahdati K., Çakmakçı R. 2019. Comprehensive biochemical insights into the seed germination of walnut under drought stress. *Scientia Horticulturae* 250: 329-343. <https://doi.org/10.1016/j.scienta.2019.02.060>
- Mozafari A.A., Ghorraishi O., Ghaderi N., Javadi T. 2016. Micropropagation of grape cultivars (*Vitis vinifera* L.) on different basal media supplemented with benzyl adenine. *Agriculturae Conspectus Scientificus* 81(3): 123-129. <https://hrca.srce.hr/178885>
- Nas M.N., Bolek Y., Sevgin N. 2010. The effects of explant and cytokinin type on regeneration of *Prunus microcarpa*. *Scientia Horticulturae* 126(2): 88-94. <https://doi.org/10.1016/j.scienta.2010.06.012>
- Panis B., Nagel M., Van den Houwe I. 2020. Challenges and prospects for the conservation of crop genetic resources in field genebanks, in *in vitro* collections and/or in liquid nitrogen. *Plants* 9(12): 1634. <https://doi.org/10.3390/plants9121634>
- Rahemi A., Gradziel T.M. 2024. Almond species classification. *The Almonds and Related Species*. Springer, Cham. [https://doi.org/10.1007/978-3-031-58938-6\\_2](https://doi.org/10.1007/978-3-031-58938-6_2)
- Rahemi A., Taghavi T., Fatahi R., Ebadi A., Hassani D., Chaparro J., Gradziel T. 2011. Seed germination and seedling establishment of some wild almond species. *African Journal of Biotechnology* 10(40): 7780-7786. <https://doi.org/10.5897/AJB10.1064>
- Renu B.A., Singh R. 2018. Optimization of micropropagation from nodal segments of apple (*Malus × domestica* Borkh) cultivars Golden Delicious and Red Fuji. *Current Journal of Applied Science and Technology* 31(3): 1-9. <https://doi.org/10.9734/CJAST/2018/45902>
- Rouhi V., Samson R., Lemeur R., Van Damme P. 2007. Photosynthetic gas exchange characteristics in three different almond species during drought stress and subsequent recovery. *Environmental and Experimental Botany* 59(2): 117-129. <https://doi.org/10.1016/j.envexpbot.2005.10.001>
- Sabbadini S., Ricci A., Limera C., Baldoni D., Capriotti L., Mezzetti B. 2019. Factors affecting the regeneration, via organogenesis, and the selection of transgenic calli in the Peach rootstock Hansen 536 (*Prunus persica* \_ *Prunus amygdalus*) to express an RNAi construct against PPV virus. *Plants* 8(6): 178. <https://doi.org/10.3390/plants8060178>
- Thakur M., Sharma V., Sharma D.P., Kumari G., Vivek M. 2016. *In Vitro* Propagation of Virus Indexed Gisela-5 (*Prunus cerasus* x *Prunus canescens*)- Clonal Cherry Rootstock. *International Journal of Crop Science and Technology* 2(2): 88-99.
- Volk G.M., Carver D.P., Irish B.M., Marek L., Frances A., Greene S., Khoury C.K., Bamberg J., del Rio A., Warburton M.L. 2023. Safeguarding plant genetic resources in the United States during global climate change. *Crop Science* 63: 2274-2296. <https://doi.org/10.1002/csc2.21003>

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